Instructor: Brent Bill, Ph.D.
Office: HPR109  Phone: 903-565-5883  Email: bbill@uttyler.edu
(The best way to contact me is to use the Canvas email system.)
Office Hours: Monday 10AM to Noon, Thursday 11AM to Noon, and by appointment

Course Overview:
Modern principles of study of cell structure, components of membranes and membrane-bound organelles, with emphasis on metabolism.

Course Objectives:
This course provides the laboratory component for Cell 3334. It is highly suggested but not required that you take this at the same time as the lecture. Biology 3134 is an intensive course with a lot of material. It will be critical that you keep up with reading, studying, and understanding the material. This lab is designed to provide you with key marketable skills that are required to be a practicing biologist and to meet National Standards suggested by the America Society of Cell Biologists. By the end of this course, student should be prepared for upper division laboratory experiences. It is expected that you will be able to find online information on genes and proteins, develop basic laboratory skills and technique, learn to work with precision equipment, and be able to write, graph, and report your findings.

Student Learning Outcomes:

<table>
<thead>
<tr>
<th>1)</th>
<th>Be able to access Biological information from the published literature and online databases (NCBI).</th>
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<tbody>
<tr>
<td>2)</td>
<td>Be able to utilize a citation manager to cite information properly.</td>
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<td>3)</td>
<td>Understand how to work safely in a biological laboratory and describe practices associated with sterile technique.</td>
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<td>4)</td>
<td>Be able to reliably use pipettes with accuracy and precision.</td>
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<td>5)</td>
<td>Understand the concepts of in vitro cell culture and perform basic cell manipulations and counting.</td>
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<td>6)</td>
<td>Understand basic concepts in microscopy (light, fluorescent, and confocal) and be able to image with bright and fluorescent microscopes.</td>
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<td>7)</td>
<td>Describe the structure and function of antibodies. Discuss how they are used specifically for immunohistochemistry and western blotting.</td>
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<tr>
<td>8)</td>
<td>Be able to utilize online databases and tools to obtain and align protein sequences. Understand the significance of shared homology.</td>
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<td>9)</td>
<td>Describe methodologies for both RNA and DNA isolation.</td>
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<td>10)</td>
<td>Understand the concept and perform Reverse Transcriptase-Polymerase Chain Reactions.</td>
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</tbody>
</table>
11) Be able to perform gel electrophoresis and understand it utilization in assessing DNA.

12) Understand how restriction enzymes cut and can be utilized for biological experiments.

13) Describe the process of PCR cloning. Explain how blue-white selection and antibiotic selection are used to obtain the proper piece of DNA.

14) Analyze DNA sequences (Sanger) via online tools.

**Required Textbooks and Readings:**

1) **BASIC CELL AND MOLECULAR BIOLOGY 4e: WHAT WE KNOW AND HOW WE FOUND OUT**
Gerald Bergstrom, 2020, *University of Wisconsin - Milwaukee*, bergstrom@uwm.edu, University of Wisconsin Milwaukee UWM Digital Commons. This text is open source and is available for free in Canvas.

2) **Labster**: An outside website that is integrated into Canvas. The College of Arts and Science is paying for your licenses ($60 per student) of Labster, so please thank Dean Gray if you see him. It will provide you with simulated pre-lab experiences, so that you will be more prepared for when you enter the real lab. It is critical that you pay attention (take notes) to the theory section and questions in the Simulations, as the material will be part of your midterm and final examinations.
   - If you need help with the Labster simulations, there is a link to the Labster support-bot in your Canvas module tab.
   - If automated responses are insufficient, it will connect you with their support staff. If you have issues, please let me know.
   - Over the course of the semester, if you have any feedback on the simulations - good or bad please email it to me, we are trying to decide if these simulations will be used for future cell laboratories, or even more broadly utilized by multiple classes in the STEM field.

3) **Dr. Bill's 2022 Cell Biology Lab Manual**: Each section will be provided in Canvas as handouts. For assignments that you need to cite the lab manual please manually add the following information into Mendeley (or other citation manager):
   - Author: Bill, Brent R.
   - Title: Cell Biology 3134 Lab Manual
   - Publication Date: 2022

**Special Course Notes:**

1) Regular Canvas access is required. carefully read all announcements. Canvas and student email should be checked **DAILY** for new announcements or messages. **All official communication should be through the Canvas email system.**
a. On Canvas you will find lectures, assignments, discussion board posts, and much more that will be essential for you as you complete this course. Dates are tentative and subject to change; therefore, it is critical to pay attention to class announcements and in lectures. Proper notice through the Canvas Announcement System will be provided if this occurs. Course grades will be maintained in Canvas. **Please note that the percentage at the end of the grade book display will not necessarily be a good indicator of your grade in the class (see below).**

2) The faculty recommends obtaining the downloadable version of Microsoft office from the University for all work. [https://www.uttler.edu/it/office365/365-students.php](https://www.uttler.edu/it/office365/365-students.php)

3) This course requires you to use the external website and a citation manager to complete homework. These are free to use but require access to the internet. Citation managers are platform specific. You can use any that work with your computers OS, and the library provides support for several versions ([https://libguides.uttler.edu/c.php?g=357640&p=2413307](https://libguides.uttler.edu/c.php?g=357640&p=2413307)).

**Assignments and point values:**

<table>
<thead>
<tr>
<th>Assignment</th>
<th>Points</th>
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</thead>
<tbody>
<tr>
<td>Midterm Exam</td>
<td>100</td>
</tr>
<tr>
<td>Final Exam</td>
<td>100</td>
</tr>
<tr>
<td>Labster</td>
<td>200</td>
</tr>
<tr>
<td>Quizzes</td>
<td>50</td>
</tr>
<tr>
<td>Benchling: Lab Notebook</td>
<td>100</td>
</tr>
<tr>
<td><strong>Total Points</strong></td>
<td><strong>550</strong></td>
</tr>
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</table>

**Grading Scale:**

A  490 or greater
B  430 to 489
C  370 to 429
D  325 to 369
F  below 324

**Late Work: No late work will be accepted!** All assignment due dates are listed. Part of learning to be a scientist is learning time management. When you are in a job or working on grants - deadlines are final! If you miss the deadline, you cannot submit; therefore, this will be good practice for your future career. If for some reason you cannot attend class or turn in an assignment, please reach out to Dr. Bill to determine if a deadline extension can be granted.

**Make-Up Exams**

You must take the exams on the scheduled dates. Please check your calendars now, so that you do not have conflicts. If an absence cannot be avoided, the professor must be notified in advance. If a student is unable to take an exam when scheduled, following appropriate documentation of the absence, the professor will arrange a make-up exam or have you take an
online version depending on the nature of the absence. Dr. Bill does not guarantee that the exam will be the same format or questions as the in-person exam.

**Acceptable Documentation:** University Note: Have your professor or coach email me a letter explaining the reason for the absence due to a prescheduled University excused absence or civil documentation: If there are other extenuating circumstances, please provide the obituary, police report, court documents, or other evidence explaining the absence.

If you are sick do not attend class or meet in person with your group. Email Dr. Bill, and we can make alternative arrangements. Zoom is available via Canvas, so that you could meet with your groups virtually.

**Attendance Policy:**
Given the hands-on nature of this course, we expect you to be in class. It is my belief that you cannot learn to work in a lab without performing the experiments. Therefore, you are required to attend:

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1 unexcused absence = - 25 points
2 unexcused absences = - 150 points, i.e. the max you could get is a C
3 unexcused absences = - 300 points, i.e. You will fail the course.
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**Documentation**
- University Note: Have your professor or coach email me a letter explaining the reason for the absence due to a prescheduled University excused absence.
- Doctors Note: If you are sick, please bring proof of your appointment, and have the doctor explain that you were indeed sick and should not or could not attend class. A note from the University COVID response team will work as well.
- Civil documentation: If there are other extenuating circumstances, please provide the obituary, police report, court documents, or other evidence explaining the absence.

**Graded Course Requirements Information TEC 51.9705 (HB 2504):**
Labster: As mentioned above, Labster is an online simulation software that will take you through virtual labs. This will prepare you by providing theory and practical experience prior to coming to the lab. It is critical to take notes as this material will be found on the exams. There will be 8 simulations.

1) Cell Culture Basics  
2) Fluorescent Microscopy  
3) Confocal Microscopy  
4) Antibodies  
5) Signal Transduction  
6) RNA Extraction  
7) Gene Regulation  
8) Molecular Cloning
**Video Quizzes:** To provide background on the subjects for the laboratory (Cytoskeleton and Transcription factors) there are 4 video lectures that have been recorded. Associated quizzes are posted in Canvas. You can take each quiz as many times as you like prior to the due date. The scores are averaged, so it is best to watch the video, take notes on it, and then take the quiz. That way you can maximize your score. This material will be on the exams.

**Lab Notebooks:** We will be using the Benchling Web-based electronic notebook (https://www.benchling.com) for class. Formats will be discussed in class. This platform will allow us to have a record of your experiences and data in the laboratory. You will be expected to keep up to date on your notebooks and data and specific analyses will be required to be uploaded. External research will be required to determine how your data should be interpreted. Periodic checks will be done by your teaching assistant and points assigned based on the quality of your notebooks and accurate representations of your data.

**Exam Policies:**
Exam questions will be drawn from the Video lectures, Labster Simulations, The Lab Manual, The Textbook, and from your practical experience gained in the lab. There will be two parts to each exam. The first part will be closed book and assess your knowledge, and the second part will be open book/computer as we will expect you to perform analyses like what you have done in class.

You are expected to follow the **University Honor Code.** All forms of cheating will be prosecuted harshly. Please note - cell phones are not allowed during the first part of the exam. Phones should be kept on the table in front of you face down during the exam. The usage of a phone during the exam is grounds for an automatic 0. The professor may video the exams to validate any instances of cheating.

**Extra Credit:**
There will be opportunities to receive up to 25 points extra credit.

1) Make a Music Video about a biological topic that we covered in class. This video should be informative about the technique as well as entertaining because we will watch these either in class or in future classes). You can work by yourself or in groups of up to 4 people. All members will receive the 25 points extra credit.

2) You must find a paper in a journal that describes a technique that we have talked about or performed in class. You should compare their methodology with that you used in class. For appropriate journals see the techniques journal links below. Please answer the following questions (5 points each):

   a) What technique did you choose to look at further?
   b) What are the benefits of their procedure, or what hurdle are they trying to overcome?
c) How is their technique different than the one we used in lab?
d) Use Mendeley to Cite the paper appropriately using the Cell Format.

**Class Expectations**

- Be Courteous and on time for classes, with assignments, and when meeting with group members.
- Silence cell phones and other electronic devices, and do not answer your phone while in class.
- Discussion is welcomed during the at any time during the lab, so please feel free to ask/answer questions, seek clarification, etc. If you need extra help, or we are pressed for time during class, please see me during office hours or ask the question in the Q&A. If I do not get questions - I will assume you understand, and I can move on - I know from experience this is not always the case and given the opportunity I will move fast.
- Please do not hesitate to ask questions! Please realize, that if submitted via email, I may not receive the question and answer it immediately; however, I will try my best. For example, if you email me Tuesday night at 11:45 P.M. the night before the exam, you may not get an answer until the next day, which may not be prior to the exam.
- If I am meeting with someone, please wait your turn. Do not just walk in and sit down. In some instances, we are discussing grades or other private matters, and it is unprofessional to interrupt.
- Due to unforeseen circumstances, I may use zoom office hours instead of in-person, If working with a student I may turn on the waiting room feature, if so please hang out until I can bring you in.
- Participate: Staying actively engaged in the lectures and labs will increase your retention. It also helps me to understand if the class is understanding the material. Therefore, throughout the lectures there are class questions. Please volunteer to answer if you think you know it. I will move around the room, so if the same people are answering do not be surprised if you get called on.
- You will be tested on your ability to work in the lab. Therefore, it is critical you are involved and being a good lab partner!

**Calendar of Topics, Readings, and Tentative Due Dates**

<table>
<thead>
<tr>
<th>Week #</th>
<th>Class Topic</th>
<th>Readings</th>
<th>Assignments Due</th>
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</table>
| Week 1 | Lab 1: Introduction to the lab (syllabus), NCBI, OMIM, Pubmed, Mendeley, Safety Tour, and Pipetting Basics. | Lab Manual Intro and Pipetting | Safety Quiz  
Safety Acknowldegement  
Contact Lens Waiver |
| Week 2 | Cell Counting and imaging. | Lab Manual Cell Counting 
CMB4: Chapter 18: The Cytoskeleton | Benchling: Entry for Lab 1. 
Labster: Cell Culture Basics, Fluorescent Microscopy  
Upload UT Tyler Biological Safety Certificate of Completion.  
Video Lecture 1-3 Quizzes |
Please note: 1/24 is the Census Date. I will be required to report attendance at this point. Please see [https://www.uttyler.edu/registrar/policies/census_date.php](https://www.uttyler.edu/registrar/policies/census_date.php) for further details regarding Census.

<table>
<thead>
<tr>
<th>Week</th>
<th>Activity</th>
<th>Lab Manual:</th>
<th>Benchling:</th>
</tr>
</thead>
<tbody>
<tr>
<td>Week 3</td>
<td>Protein Extraction and Quantitation</td>
<td>Protein Extraction</td>
<td>Entry for Cell Counting and Staining</td>
</tr>
<tr>
<td>Week 4</td>
<td>SDS-PAGE and Western Blotting Started</td>
<td>SDS PAGE</td>
<td>Antibodies</td>
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<td></td>
<td></td>
<td></td>
<td>Entry for Protein Extraction and Quantitation</td>
</tr>
<tr>
<td>Week 5</td>
<td>Western Blot Development, Immunohistochemistry</td>
<td>Western Blot and IHC</td>
<td>Confocal Microscopy, Signal Transduction, Entry SDS-PAGE and Western.</td>
</tr>
</tbody>
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**W 2/9**  
Start of Darwin Week!  
**R 2/10**  
Darwin Day at TJC!  
**F 2/11**  
Darwin Day at UT Tyler!

<table>
<thead>
<tr>
<th>Week 6</th>
<th>Bioinformatics</th>
<th>Lab Manual Bioinformatics</th>
<th>Entry for Western Blot and IHC.</th>
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<tbody>
<tr>
<td>Week 7</td>
<td>No Class— Study week. Dr Bill and TA are at TAS Conference (Thursday and Friday)</td>
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<td>Entry for Bioinformatics.</td>
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<tr>
<td>Week 8</td>
<td>Midterm Exam</td>
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Please note: Tuesday, March 1 is the Last Day to Register for Spring Graduation. Spring Break! Officially March 7-12.

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<tr>
<th>Week 10</th>
<th>RNA Isolation</th>
<th>Lab Manual: RNA Isolation CMB4e: Chapter 12.5, 12.5.1, 15.2</th>
<th>Labster: RNA Isolation Video Lecture Quiz 4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Week 11</td>
<td>RT-PCR</td>
<td>Lab Manual RT-PCR CMB4e: Chapter 15.5</td>
<td>Labster: Gene Regulation</td>
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<td></td>
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<td></td>
<td>Benchling: Entry for RNA Isolation</td>
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</table>

Please note that Monday, March 28, is the last day to withdraw from classes (W).

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<tr>
<th>Week 12</th>
<th>Plasmid Minipreps, Restriction Digests, and Agarose Gel Electrophoresis</th>
<th>Lab Manual: Minipreps, Digests, and Gels.</th>
<th>Labster: Molecular Cloning</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Benchling: Entry for RT-PCR</td>
</tr>
<tr>
<td>Week 14</td>
<td>No Class: Study week</td>
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<td>Benchling: Entry for Sequence analysis and primer Design.</td>
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<tr>
<td>Week 15</td>
<td>Final Exam</td>
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Spring Commencement: May 7