THE UNIVERSITY OF TEXAS AT TYLER
INSTITUTIONAL BIOSAFETY COMMITTEE
May 20, 2026

MEMBERS' PRESENT:

Dr. Galina Florova – IBC Chair – Professor Cellular and Molecular Biology
Dr. Buka Samten – IBC Member - Associate Professor of Microbiology
Dr. Wei-Chin Ho – IBC Member – Assistant Professor of Biology
Dr. Dustin Patterson – IBC Member - Professor of Chemistry & Biochemistry
Dr. Hua Tang – IBC Member – Professor of Biochemistry
Mr. Christopher Frydenlund - Senior Manager Research Compliance – Biosafety Officer
Ms. Beth R. Filla – Local Non-affiliated member - Bachelor of Science in Agricultural Development
Mr. Michael Medford - Local Non-affiliated member - P.E., CPESC

OTHERS PRESENT:
Recorder Secretary

MEMBERS ABSENT:
Dr. Wei-Chin Ho joined the meeting at 1:10 p.m.

OLD BUSINESS:
The committee continues to meet via TEAMS.

I. CALL TO ORDER:
The meeting was called to order at 1:00 p.m.

II. APPROVAL OF MINUTES:
Minutes from March 20, 2026,IBC meeting were approved without changes.
The committee reviewed the minutes, and a motion was made by Dr. Dustin Patterson to approve the minutes. Dr. Buka Samten seconded the motion.

APPROVE: 8 ABSTAIN: 0 OPPOSE: 0

III. REVIEW OF IORRC MINUTES:
The minutes from February 18, 2026, meeting were reviewed by the IBC.

IV. CHAIRMAN'S REPORT:
None

V. OLD BUSINESS:
None

VI. NEW BUSINESS:

Protocol and Addendum for review

Recombinant DNA protocols

rDNA Protocol - 268

Protocol	(PI)	Project Title	BSL	ABSL	Campus
268	**Dr. L.V. Rao**	Elucidation of Hemostatic Thrombotic and Inflammatory Mechanisms in Health and Disease Conditions.	**BSL2 & BSL3**		**North Campus**

Project overview

The primary focus of our studies is to understand the mechanisms that regulate bleeding, clotting, and inflammation in health and disease. Another aim is to understand how blood clotting and fibrinolytic proteins contribute to various diseases, including inflammation, cardiovascular diseases, cancer, and infection. To understand the structure and function of various clotting proteins, we will construct recombinant DNA encoding normal or mutant clotting proteins. These recombinant DNAs will be constructed in either *Escherichia coli* (*E. coli*), adenovirus, retrovirus, or lentivirus. We will introduce the recombinant DNA into different cell types, such as endothelial cells (the cells that line blood vessel walls), muscle cells, kidney cells, or cancer cells. We will also use various siRNAs and microRNAs to suppress the expression of specific genes. We will use silico, cell, and animal model systems to investigate the effects of normal and mutant proteins generated by recombinant DNA technology on blood coagulation, inflammation, and various disease processes. Bacterial and viral vectors enclosing recombinant DNA relevant to our research will be administered to animals to understand the role of various clotting, inflammatory, signaling, and other molecules in pathophysiological processes. For animal studies, we may generate knock-out or transgenic mice for mediators that regulate hemostasis, thrombosis, inflammation, and other disease processes.

Applicable NIH Guidelines: Section II-A-3, III, and IV-B-2-b

Major points of discussion:

- Agent characteristics (e.g. virulence, pathogenicity, environmental stability)
- Types of manipulations planned.
- Source(s) of the nucleic sequences (e.g., species)
- Nature of the nucleic acid sequences (e.g., structural gene, oncogene)
- Host(s) and vector(s) to be used
- Whether an attempt will be made to obtain expression of a transgene, and if so, the function of the protein that will be produced.

The following will be communicated to the PI:

Page 2 **Experimental Plan in Lay Terms -** Editorial changes are required.

Page 5 **A. Host**

- Please clarify if produced strains would change pathogenicity.
- Please specify the pathogenicity of *E. coli* strains.
- Please clarify if every strain of the mouse will be used as a host for the future modifications.

Page 6 **B. Vector –** Editorial change is required on second line under promoter tab (T7Lab)

Page 8 **C. Gene –** Please clarify who will be the investigators that you intend to use as collaborators.

F. Risks to Human

- Clarify what work will be conducted under BSL2 and BSL3 conditions.
- State that you will follow the BSL2 and BSL3 manuals.

Page 9C. Animals

- Editorial changes are required.

The committee reviewed the protocol, and a motion was made by Dr. Buka Samten to approve the protocol pending corrections. Dr. Dustin Patterson seconded the motion.

APPROVE: 8 ABSTAIN: 0 OPPOSE: 0

Recombinant DNA protocols

rDNA Protocol – 262 Addendum 1

Protocol	(PI)	Project Title	BSL	ABSL	Campus
262-1	**Dr. C. Abdullah**	Novel Molecular Mechanisms of Lipid Metabolism.	**BSL1 & BSL2**		**Main Campus**

Project overview:

Committee requested Dr. Abdullah to focus on the objective of the addendum and the corresponding protocol and describe those accordingly.

Applicable NIH Guidelines: Section II-A-3, III, and IV-B-2-b

Major points of discussion:

- Agent characteristics (e.g. virulence, pathogenicity, environmental stability)
- Types of manipulations planned.
- Source(s) of the nucleic sequences (e.g., species)
- Nature of the nucleic acid sequences (e.g., structural gene, oncogene)
- Host(s) and vector(s) to be used
- Whether an attempt will be made to obtain expression of a transgene, and if so, the function of the protein that will be produced.

The following will be communicated to the PI:

The institutional Biosafety Committee was very concerned about the clarity and consistency of information provided and is not ready to approve this addendum at this time. The IBC members agree that a new protocol submission is required because the addendum introduces multiple genes that extends it beyond the scope of the original protocol, and does not provide enough detail about the vectors, hosts, or associated risks to humans.

The committee reviewed the protocol, and a motion was made by Dr. Dustin Patterson to table the protocol. Dr. Hua Tang seconded the motion.

APPROVE: 8 ABSTAIN: 0 OPPOSE: 0

Recombinant DNA protocols

rDNA Protocol – 257 Addendum 2

Protocol	(PI)	Project Title	BSL	ABSL	Campus
257-2	**Dr. M. Abdelaziz**	Generating Gene Deletions in Staphylococcus aureus	**BSL2**		**Main Campus**

Project overview:

The objective of this protocol is to study the TCS crosstalk in *S. aureus* signaling network.

Addendum 2 – Adding a second host strain that will harbor all the proposed gene deletions under section 6A.

Applicable NIH Guidelines: Section II-A-3, III, and IV-B-2-b

Major points of discussion:

- Agent characteristics (e.g. virulence, pathogenicity, environmental stability)
- Types of manipulations planned.
- Source(s) of the nucleic sequences (e.g., species)
- Nature of the nucleic acid sequences (e.g., structural gene, oncogene)
- Host(s) and vector(s) to be used
- Whether an attempt will be made to obtain expression of a transgene, and if so, the function of the protein that will be produced.

The following will be communicated to the PI:

Page 1 Editorial changes required.

The committee reviewed the protocol, and a motion was made by Dr. Buka Samten to approve the protocol pending corrections. Dr. Wei-Chin Ho seconded the motions.

APPROVE: 8 ABSTAIN: 0 OPPOSE: 0

Recombinant DNA protocols

rDNA Protocol – 258- Addendum 2

Protocol	(PI)	Project Title	BSL	ABSL	Campus
258-2	**Dr. M. Abdelaziz**	VraS Mutant by Recombineering and CRISPR/Cas9 Counterselection.	**BSL 2**		**Main Campus**

Project overview:

The objective of this protocol is to assess the changes in antibiotic MIC that occur after introducing nine targeted single point mutations to the genome of *S. aureus*.

Addendum 2 – Adding a second *S. aureus* host strain: SH1000

Applicable NIH Guidelines: Section II-A-3, III, and IV-B-2-b

Major points of discussion:

- Agent characteristics (e.g. virulence, pathogenicity, environmental stability)
- Types of manipulations planned.
- Source(s) of the nucleic sequences (e.g., species)
- Nature of the nucleic acid sequences (e.g., structural gene, oncogene)
- Host(s) and vector(s) to be used
- Whether an attempt will be made to obtain expression of a transgene, and if so, the function of the protein that will be produced.

The following will be communicated to the PI:

Page 2

- Please clarify absence of antibiotic resistance stated for the strain(s) proposed for the study
- Please add a statement regarding the risks of humans, you may reuse the one that is already in a protocol if it is appropriate.

The committee reviewed the protocol, and a motion was made by Dr. Buka Samten to approve the protocol pending corrections. Dr. Dustin Patterson seconded the motion.

APPROVE: 8 ABSTAIN: 0 OPPOSE: 0

VII. ADJOURNMENT:

The meeting was adjourned at 2:54 p.m.

DocuSigned by:
Galina Florova 7/27/2026
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Galina Florova, Ph.D. – Chair
Institutional Biosafety Committee

Certificate Of Completion

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In Person Signer Events	Signature	Timestamp

Editor Delivery Events	Status	Timestamp

Agent Delivery Events	Status	Timestamp

Intermediary Delivery Events	Status	Timestamp

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Witness Events	Signature	Timestamp

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Envelope Summary Events	Status	Timestamps
Envelope Sent	Hashed/Encrypted	7/23/2026 9:41:29 AM
Certified Delivered	Security Checked	7/27/2026 12:24:45 PM
Signing Complete	Security Checked	7/27/2026 12:25:12 PM
Completed	Security Checked	7/27/2026 12:25:12 PM

Payment Events	Status	Timestamps